

Diagnostics for genotypes from low-depth sequencing



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Introduction

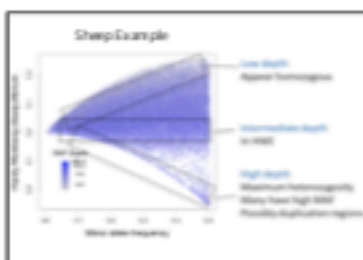
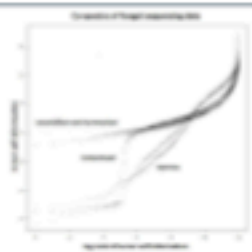
- Sequencing is increasingly being used for genotyping. Costs can be reduced by sequencing a representative fraction of the genome at low depth.
- Low depth genotypes may be uncertain due to low read depth; statistical methods are being developed to account for this uncertainty.
- Diagnostics are important for checking the laboratory and computational workflows.
- We present some depth-aware diagnostics that should be used in conjunction with knowledge about the samples.
- These diagnostics are available in scripts in the AgResearch GitHub repository (<https://github.com/AgResearch>).

Diagnostic Tools



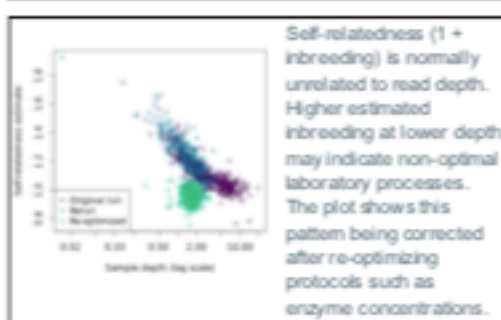
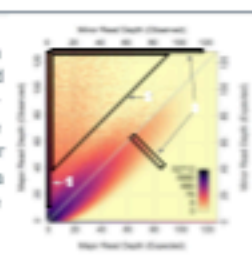
Graphical display of a measure of sequencing success against the physical arrangement of samples during lab processing. Here the sample depth (mean number of reads over detected SNPs) is displayed by 384-well plate layout and shows typical variation. Negative controls are marked with X. Any patterns in sequencing success may reflect sample mishandling.

Genome sequences display characteristic signatures for species and contaminants. McCulloch et al. (2018) show how a k-mer ziplan plot of self-information in k-mers against their rank can be used to diagnose contaminated and repetitive samples.



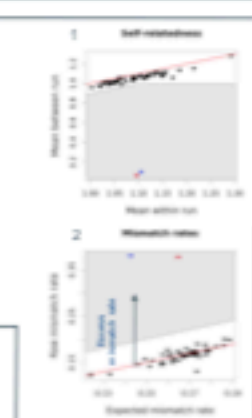
A fin plot (Dodds et al., 2015) of Hardy-Weinberg disequilibrium (HWD) against minor allele frequency (MAF) for each SNP can help detect repeated regions erroneously assigned to the same SNP position—they have low HWD for their MAF and relatively high depth. The plot may also show that many SNPs with high apparent HWD have low depth so heterozygotes may appear homozygous.

The comet plot (Bliton, 2020) shows the distribution of allele count pairs mirrored against that expected under binomial sampling. Example insights: 1) sequencing error rate (homozygotes), 2) extra-binomial sampling, 3) allele count limit in the software used.



Self-relatedness (1 + inbreeding) is normally unrelated to read depth. Higher estimated inbreeding at lower depth may indicate non-optimal laboratory processes. The plot shows this pattern being corrected after re-optimizing protocols such as enzyme concentrations.

Sometimes an individual is re-genotyped. Comparing the low-depth genotype calls can be problematic as one allele may not be observed. Two alternative approaches are: 1) compare relatedness estimates between the results to the mean self-relatedness (expected to be similar for the same individual) and 2) calculate the excess mismatch rate, the difference between the observed (raw) mismatch rate and the mismatch rate that is expected given the read depths. This is analogous to the parentage methods in Dodds et al. (2019). Both plots highlight the same two incorrectly labelled individuals (♦, ♦).



Conclusion

Depth-aware diagnostics help QC-check genotypes from low depth sequencing.

References

- Bliton T (2020) PhD Thesis, University of Otago
- Dodds K et al. (2015) *Genetics* 191: 1017-1024
- Dodds K et al. (2019) *Genetics* 207: 1017-1024
- McCulloch K et al. (2018) *Genetics* 200: 1017-1024

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