



WHAT IF ALL VARIANTS ARE RARE?

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ROTHAMSTED
RESEARCH

INTRODUCTION



Figure 1. Wheat TILLING plot in the wheat field. Scale bar is 100 m (Lyrer et al. 2017).

- TILLING ($T_{\text{rate}} \approx \text{mut} \cdot L_{\text{seq}} \cdot L_{\text{pop}}$ In G_{genome}) is a method used to create a **mutagenised** population allowing the identification of point mutations within genes of interest (Krasileva et al. 2017).
- The relative contribution of (**ultra**) **rare variants** to quantitative genetic variation is largely unclear, despite recent advances in genomics. One reason for this is that natural populations tend to violate the assumptions of **genome-wide association** and **genomic heritability** models.
- Field-based phenotyping tools have been used to identify genes/QTL throughout the crop growth cycle for **dynamic phenotypes** (Lyrer et al. 2020).

MAIN OBJECTIVES

The goals of this study were to:

- perform **gene-based association analysis** using collapsing models into **non-synonymous**, **synonymous**, and **non-coding mutations** sets,
- partition the variance of (a) mutation types over the course of the growing season, (b) single genes, and (c) chromosomes.

QUANTITATIVE GENETICS PIPELINE



MATERIALS AND METHODS

- We performed quantitative genomic analyses (see pipeline) in an experimental TILLING population of hexaploid wheat cv. Cadizna (~1145 lines) in which all chemically induced mutations were either rare or unique, starting with a (mostly) uniform genomic background.
- The field trials were conducted in John Innes Centre (JC) and at the Field Scanner phenotyping platform at Rothamsted Research, Harpenden, UK.
- Using custom re-sequencing data, we partitioned the complete set of ~7M mutations (~77k genes) into non-synonymous, synonymous, and non-coding sets.
- We conducted **gene-based analysis** (collapsing models) and variance component analysis for three phenotypic traits (i.e. plant height, heading/flowering time, and senescence).

GENE SCANNING AND VARIANCE PARTITIONING

- Gene scanning of variants revealed **association signals** coming mostly from **non-synonymous** and **no non-synonymous substitutions** for plant height (Fig. 3).
- Breaking down the genetic variance of **chromosomes** and **single genes** revealed insights of the contribution of **ultra-rare variants** to phenotypic traits (Fig. 3).

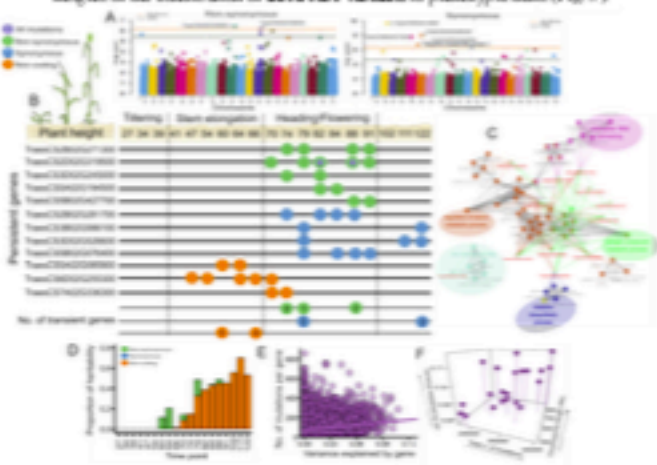


Figure 3. Overview of the quantitative genetic analysis of plant height in the wheat TILLING population. (A) Manhattan plot of association signals for plant height in the wheat TILLING population. (B) Bar chart showing the number of variant genes for different mutation types. (C) Network diagram showing gene-gene interactions. (D) Bar chart showing the number of variant genes per chromosome. (E) Scatter plot showing the number of variant genes per gene. (F) Scatter plot showing the number of variant genes per chromosome.

ULTRA-RARE VARIANTS IN THE WHEAT TILLING POPULATION

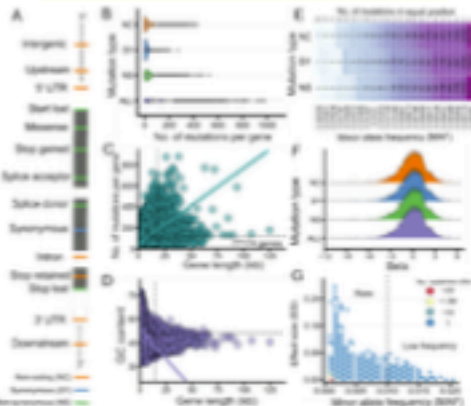


Figure 5. Mutation variant summary in the wheat TILLING population. (A) Chromosome diagram of gene scanning. (B) Scatter plot showing the number of variant genes per gene. (C) Scatter plot showing the number of variant genes per chromosome. (D) Scatter plot showing the number of variant genes per gene. (E) Bar chart showing the number of variant genes per chromosome. (F) Scatter plot showing the number of variant genes per gene. (G) Bar chart showing the number of variant genes per chromosome.

CONCLUSION

This approach will lead to the identification of **candidate genes** across a range of phenotypic traits and contribute to the development of novel analytical methodology, ultimately enhancing our understanding of the **importance** of (**ultra**) rare variants in both natural and experimental populations.

REFERENCES

- Krasileva, K.V. et al. (2017) A mutagenesis pipeline to generate wheat populations for the study of natural variation. *Plant Science*, 258, 1-10.
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